Pleuromutilin Antibiotic

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SB-275833 Altabax[®]

(3aS,4R,5S,6S,8R,9R,9aR,10R)-2-(exo-8-Methyl-8-azabicyclo[3.2.1]octan-3-ylsulfanyl)acetic acid 5-hydroxy-4,6,9,10-tetramethyl-1-oxo-6-vinylperhydro-3a,9-propanocyclopentacycloocten-8-yl ester

Mutilin 14-(exo-8-methyl-8-azabicyclo[3.2.1]oct-3-ylsulfanyl)acetate

C₃₀H₄₇NO₄S Mol wt: 517.7645

CAS: 224452-66-8

EN: 276526

Abstract

Due to the emergence of multidrug-resistant microorganisms, research efforts are focusing on new classes of antibacterials with different mechanisms of action from those in clinical use. One such agent is retapamulin, a pleuromutilin derivative with a unique mechanism of action. Currently under development for the topical treatment of skin infections, retapamulin has shown no target-specific cross-resistance to commonly used antibiotics. In a number of in vitro studies, retapamulin demonstrated activity that was equivalent or superior to other common antibiotics against clinical isolates from skin and skin structure infections. The organisms susceptible to retapamulin include: Streptococcus pyogenes, Streptococcus agalactiae, β-hemolytic streptococci, viridans streptococci, Staphylococcus aureus, coagulase-negative staphylococci (including Staphylococcus epidermidis), Propionibacterium spp. (including Propionibacterium acnes), Prevotella spp., Porphyromonas spp. and Fusobacterium spp. However, retapamulin showed minimal or no activity against entero-Gram-negative bacilli. Importantly, retapamulin maintains its activity against organisms that are resistant to a number of antimicrobial agents, including methicillin, erythromycin, fusidic acid, mupirocin, azithromycin and levofloxacin. Evidence from multiplestep and single-step studies also suggests that retapamulin has a low potential to select for resistant mutants in S. pyogenes and S. aureus.

Synthesis

Reaction of the natural antibiotic pleuromutilin (I) with methane sulfonyl chloride by means of either TEA or DIEA in either $\mathrm{CH_2Cl_2}$ or methyl isobutyl ketone provides the corresponding mesylate derivative (II) (1), which is submitted to nucleophilic substitution with exo-8-methyl-8-azabicyclo[3.2.1]octan-3-thiol (III) by means of either tetrabutylammonium hydrogensulfate in $\mathrm{CH_2Cl_2/H_2O}$ and NaOH at pH 12.5 (1) or potassium tert-butoxide in ethanol (2). Scheme 1.

The azabicyclic thiol derivative (III) can be prepared by different ways:

- 1) Mitsunobu reaction of *endo*-8-methyl-8-azabicy-clo[3.2.1]octan-3-ol (IV, tropine) with triphenylphosphine, diisopropyl azodicarboxylate and thioacetic acid in dry THF (2). Scheme 1.
- 2) Treatment of endo-8-methyl-8-azabicyclo[3.2.1]-octan-3-ol (IV, tropine) with methane sulfonyl chloride and either TEA or DIEA in $\mathrm{CH_2Cl_2}$ yields tropine-3-mesylate (V), which after nucleophilic substitution with sodium ethyl dithiocarbonate in toluene affords the intermediate (VI). Finally, the dithiocarbonate derivative (VI) is hydrolized by means of NaOH and then treated with HCI (1). Scheme 1.

Background

Pleuromutilin was first isolated in 1951 from *Pleurotus mutilus*, an edible mushroom. Nearly two decades later, this compound was discovered to have excellent activity against *Mycoplasma* spp. Tiamulin and valnemulin are semisynthetic derivatives of pleuromutilin employed for veterinary use, and a third analogue, azamulin, was studied as a potential antibiotic for use in humans. When the pleuromutilins were first investigated, most research was focused on the development of β -lactam, macrolide and quinolone antibacterial agents. However, more recently, due to the emergence of multidrug-resistant pathogens,

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efforts are being made to develop new classes of antimicrobials with novel modes of action and activity against resistant organisms. This strategy has led to the reassessment of previously discovered antibacterial agents that have not yet been used in humans. Pleuromutilins display a unique mode of action, inhibiting bacterial protein synthesis by specifically targeting the large subunit of the bacterial ribosome (3). Retapamulin (SB-275833) is a semisynthetic member of the pleuromutilin class of antibiotics, developed for the topical treatment of uncomplicated skin and skin structure infections (SSSIs) due to susceptible strains of *Staphylococcus aureus* or *Streptococcus pyogenes*, particularly impetigo (4-6).

Preclinical Pharmacology

In a recent study characterizing the interactions of retapamulin with bacterial ribosomes, results indicated that retapamulin is a selective and potent inhibitor of bacterial protein synthesis. This compound had IC $_{50}$ values of 0.33 and > 100 μ M in a bacterial coupled transcription-translation assay and a eukaryotic translation assay system, respectively. In addition, retapamulin showed extremely high binding affinity (K $_{\rm d}$ approx. 3 nM) for Escherichia coli and S. aureus ribosomes. The compound also potently inhibited bacterial ribosomal peptidyltransferase (IC $_{50}$ = 26 nM). Overall, data from this study indicate that the retapamulin binding site on the 50S ribosomal subunit involves nucleotides in the region of the ribosomal P-site and peptidyltransferase center. Retapamulin inhibited fmet-tRNA binding to the P-site of

 $E.\ coli$ ribosomes in vitro (IC $_{50}$ = 29 nM), a finding which suggests that the compound may have an effect on translation initiation. Furthermore, retapamulin interacts with domain V of 23S rRNA in a manner that is different from the macrolides and ketolides (7).

The in vitro activity of retapamulin was examined using organisms isolated from SSSIs. The MIC_{90} values for retapamulin against nearly 4,000 clinical isolates of staphylococci, β-hemolytic streptococci (including S. pyogenes and Streptococcus agalactiae) and viridans streptococci were determined. The MIC₉₀ values against clinical isolates of S. aureus were 0.12 μg/ml for retapamulin and 16, > 128, 32, 8-16, > 32, 2, > 32, 1, 2, 2-4, 4, 64 or more, 32 and 16-32 µg/ml, respectively, for amoxicillin/clavulanic acid, bacitracin, ceftriaxone, cephalothin, clindamycin, cloxacillin, erythromycin, fusidic acid, gentamicin, linezolid, mupirocin, neomycin, penicillin and tetracycline. Against isolates of coagulase-negative staphylococci, including S. epidermidis, the MIC_{on} values were 0.06-0.12, 8, > 128, 32-64, 4, > 32, 16, > 32, 4-8, 16, 2. > 256. 16-32. 16 and $32 \mu g/ml$ or more for the respective agents. Against clinical isolates of S. pyogenes, the MIC_{90} value for retapamulin was 0.03-0.06 $\mu g/ml$, as compared to MIC_{90} values of 0.12, 32-64, 0.06-0.12, 0.25, 0.12-0.25, 0.5, 2, 8, 16, 1, 0.5, 64, 0.03-0.06 and 16-32 μg/ml, respectively, for amoxicillin/clavulanic acid, bacitracin, ceftriaxone, cephalothin, clindamycin, cloxacillin, erythromycin, fusidic acid, gentamicin, linezolid, mupirocin, neomycin, penicillin and tetracycline. Similarly, for clinical isolates of S. agalactiae, the MIC₉₀ values were $0.06 \mu g/ml$ for retapamulin and 0.25, 128, 0.12, 0.5,

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4-32, 2, 8-32, 16, 32, 1, 1, 64 or more, 0.12 and > 32 μg/ml, respectively, for the other agents. Against strains of viridans streptococci, the MIC $_{90}$ value for retapamulin was 0.25 μg/ml, as compared to 1, 128, 1, 1-2, 4-16, 8, 16-> 32, 32, 16, 1, 2, 64 or more, 1 and 32 μg/ml or more, respectively, for the comparators. Against macrolideresistant strains of S. pyogenes, retapamulin showed an MIC $_{90}$ value of 0.06 μg/ml, as compared to an MIC $_{90}$ value of 0.03 μg/ml for macrolide-susceptible strains. Retapamulin retained its *in vitro* activity against strains resistant to methicillin, erythromycin, mupirocin or fusidic acid (8-13).

Retapamulin demonstrated excellent antibacterial activity against 392 clinical isolates of Gram-positive and Gram-negative organisms. In this study, retapamulin had MIC₉₀ values of 0.125, 0.125, 0.125, 0.016, 0.125, 0.03, 0.125, 2 and 0.03 μg/ml, respectively, against S. aureus S. epidermidis (40 isolates), Staphylococcus saprophyticus (28 isolates), S. pyogenes (101 isolates), Streptococcus pneumoniae (26 isolates), S. agalactiae (27 isolates), Streptococcus viridans (23 isolates), Haemophilus influenzae (27 isolates) and Moraxella catarrhalis (20 isolates). For comparison, mupirocin had MIC_{90} values of 0.5, > 16, 8, 0.5, 4, 1, 2, 0.125 and 2 μg/ml, respectively, against the same organisms; fusidic acid gave respective MIC₉₀ values of 0.5, 0.25, 8, 16, 32, 16, 32, 16 and 0.06 $\mu g/ml$; bacitracin showed MIC_{on} values of 2 μg/ml for S. pyogenes, 4 μg/ml for *M. catarrhalis* and > 16 μg/ml for all other organisms tested; amoxicillin yielded MIC₉₀ values of 64, 0.5, 0.5, 0.016 or less, 2, 0.06, 8, > 64 and 8 μ g/ml against S. aureus, S. epidermidis, S. saprophyticus, S. pyogenes, S. pneumoniae, S. agalactiae, S. viridans, H. influenzae and M. catarrhalis, respectively; azithromycin gave respective MIC_{90} values of > 64, > 64, > 64, 4, > 8, 2, > 8, 2 and 0.03 μg/ml; levofloxacin had MIC₉₀ values of 16, 8, 0.5, 2, 1, 1, 2, 0.016 and 0.03 μ g/ml against the respective microorganisms; and cefaclor showed MIC_{on} values of > 64, 1, 4, 0.25, > 8, 2 > 8, > 16 and 2 μ g/ml, respectively (14).

The in vitro activity of retapamulin was tested against 1,690 Gram-positive clinical isolates commonly associated with SSSIs collected between 1998 and 2004 in the U.S. and Europe. The MIC₉₀ values for retapamulin were 0.25, 0.06, 0.12, 0.25, 0.5, 0.06, 128, 0.5 and $0.25 \mu g/ml$, respectively, against S. aureus (500 isolates), S. pyogenes (500 isolates), S. epidermidis (100 isolates), S. saprophyticus (100 isolates), viridans streptococci (101 isolates), S. agalactiae (100 isolates), Bacillus spp. (102 isolates), Corynebacterium spp. (101 isolates) and Micrococcus spp. (86 isolates). Retapamulin exhibited a better profile of in vitro activity than bacitracin, cephalothin, ciprofloxacin, erythromycin, gentamicin, mupirocin, oxacillin and penicillin, and maintained good activity against methicillin-resistant, vancomycin-intermediate and vancomycin-resistant strains (MIC = 0.06-0.5 μg/ml)

The *in vitro* activities of retapamulin, erythromycin, fusidic acid and mupirocin were determined against 503

isolates of *S. pyogenes*, 130 isolates of *S. agalactiae* and 105 isolates of viridans streptococci. The MIC_{90} values found for retapamulin were 0.03, 0.06 and 0.25 μ g/ml, respectively, vs. respective values for erythromycin of 16, 16 and 128 μ g/ml or more, for fusidic acid of 8, 16 and 32 μ g/ml and for mupirocin of 0.12, 1 and 1 μ g/ml (16).

The antibacterial activities of retapamulin and comparators were evaluated against a total of 604 recent clinical isolates of S. aureus (281 strains), coagulase-negative staphylococci (232 strains) and S. pyogenes (91 strains), including oxacillin-, mupirocin- and erythromycinresistant strains. Retapamulin showed an MIC₉₀ value of 0.12 μg/ml against all drug-resistant strains of S. aureus tested, as well as against wild-type susceptible S. aureus. For comparators, the MIC₉₀ values against susceptible S. aureus isolates were 1, > 4, 0.25 or less, 0.12 or less, 0.5, 2, 0.5 and > 2 μg/ml, respectively, for tiamulin, bacitracin, cephalothin, clindamycin, gentamicin, linezolid, ofloxacin and penicillin. Against erythromycin-resistant strains of S. aureus, the MIC₉₀ values for the respective agents were 1, > 4, > 32, > 100, 30, 2, > 30 and > $2 \mu g/ml$. The MIC_{on} values against mupirocin-resistant S. aureus isolates were 1, > 4, > 32, > 100, 30, 2, > 30 and $> 2 \mu g/ml$ for the respective comparators. Against oxacillin-resistant strains of S. aureus, the MIC_{90} values were 1, > 4, > 32, > 100, 30, 2, > 30 and > 2 μ g/ml, respectively. The MIC_{on} value of retapamulin against coagulase-negative staphylococci was 0.06 µg/ml for both oxacillin- and mupirocin-resistant strains, as well as for wild-type susceptible isolates. The MIC_{on} values against susceptible coagulase-negative staphylococci isolates were 0.5, > 4, 0.25 or less, 0.12 or less, 0.25, 2, 0.5 and $> 2 \mu g/ml$, respectively, for tiamulin, bacitracin, cephalothin, clindamycin, gentamicin, linezolid, ofloxacin and penicillin. Against mupirocin-resistant strains of coagulase-negative staphylococci, the MIC90 values were 0.5, > 4, 2, > 100, 30, 1, > 30 and $> 2 \mu g/ml$ for the respective agents. The MIC₉₀ values against oxacillin-resistant coagulase-negative staphylococci were 0.5, > 4, 4, > 100, 30, 1, > 30 and $> 2 \mu g/ml$, respectively. Retapamulin gave an MIC_{90} value of $0.03 \,\mu g/ml$ or less against erythromycin-resistant S. pyogenes, as compared to MIC_{on} values of 0.25 or less, 2 or less, > 4, 0.25 or less, > 100, 8, 1, 1, 0.25 or less and 0.03 μ g/ml or less, respectively, for tiamulin, mupirocin, bacitracin, cephalothin, clindamycin, gentamicin, linezolid, ofloxacin, oxacillin and penicillin (17).

Another recent study investigated the *in vitro* activity of retapamulin against 141 clinical isolates of *Propionibacterium* spp., including 7 multiresistant strains. Against 117 strains of *Propionibacterium acnes*, the MIC $_{90}$ values were 0.25, 0.25, 0.06, 1 and 1 μ g/ml for retapamulin, clindamycin, erythromycin, tetracycline and trimethoprim/sulfamethoxazole, respectively. In addition, retapamulin had an MIC of 1 μ g/ml or less against *P. acnes* strains resistant to comparator drugs, including 5 strains with erythromycin MIC values of > 128 μ g/ml, 3 strains with tetracycline MIC values of 8 μ g/ml or more and 3 strains with clindamycin MIC values of 4 μ g/ml or more. When tested against 12 strains of

Propionibacterium avidum, retapamulin gave an MIC $_{90}$ value of 0.015 μg/ml, as compared to values of 0.06, 0.125, 4 and > 8 μg/ml, respectively, for clindamycin, erythromycin, tetracycline and trimethoprim/sulfamethoxazole. The MIC $_{90}$ values against *Propionibacterium granulosum* were 0.015 or less, 0.03, 0.125, 2 and > 8 μg/ml for retapamulin, clindamycin, erythromycin, tetracycline and trimethoprim/sulfamethoxazole, respectively (18, 19).

The in vitro activity of retapamulin was determined against 232 isolates of anaerobic organisms collected in 2003 and 2004 from human skin samples. Against 25 strains of Bacteroides fragilis, the MIC₉₀ values were 64, > 64, 16, 64, 4, > 256 and 4 μ g/ml, respectively, for retapamulin, amoxicillin, amoxicillin/clavulanic acid, ceftriaxone, imipenem, clindamycin and metronidazole. The MIC₉₀ values against 10 strains of Bacteroides thetaiotaomicron were > 64, > 64, 2, > 128, 2, > 256 and $2 \mu g/ml$ for the respective agents. Against 17 strains of other species in the *B. fragilis* group, the MIC_{90} values were 32, > 64, 16, > 128, 1, > 256 and 4 μ g/ml, respectively. When tested against 33 strains of Prevotella spp., the MICon value of retapamulin was 0.03 μg/ml, as compared to 0.125, 0.06 or less, 2, 0.03 or less, 0.06 or less and 0.5 μg/ml, respectively, for amoxicillin, amoxicillin/clavulanic acid, ceftriaxone, imipenem, clindamycin and metronidazole. Against 29 strains of Porphyromonas spp., the MIC_{on} values were 0.015, 0.06, 0.06, 0.03, 0.03, 0.06 and 0.06 µg/ml or less, respectively, for retapamulin, amoxicillin, amoxicillin/clavulanic acid, ceftriaxone, imipenem, clindamycin and metronidazole, and the MIC_{on} values determined against 25 strains of Fusobacterium spp. were 2, 16, 0.5, 128, 0.5, 4 and 0.5 μg/ml for the respective agents. Testing against 14 strains of Clostridium perfringens yielded MIC_{90} values of 1, 0.06 or less, 0.06 or less, 4, 0.5, 0.5 and 1 µg/ml, respectively, and the corresponding MIC₉₀ values against 15 strains of other Clostridium spp. were 64, 2, 2, 64, 2, 256 and 0.5 μg/ml. Against 25 strains of P. acnes, the respective agents yielded MIC₉₀ values of 0.25, 0.25, 0.25, 0.5, 0.125, 0.25 and > 64 μg/ml. Retapamulin also displayed good activity against Gram-positive anaerobic cocci, with an MIC_{qn} of 1 μ g/ml compared to 0.25, 0.25, 8, 0.25, 32 and 2 μ g/ml, respectively, for amoxicillin, amoxicillin/clavulanic acid, ceftriaxone, imipenem, clindamycin and metronidazole (20).

Another study involving 226 strains of Gram-negative and Gram-positive anaerobic bacteria collected from human skin samples compared the *in vitro* activity of retapamulin with clindamycin, metronidazole, mupirocin, neomycin and tetracycline. The MIC $_{90}$ values were 64, > 128, 2, > 128, > 128 and 128 μ g/ml, respectively, against 35 strains belonging to the *B. fragilis* species group, 4, 0.5, 8, 1, 4 and 16 μ g/ml, respectively, against 23 *Bacteroides* spp. strains, 1, 0.125, 1, 0.06, > 128 and 2 μ g/ml, respectively, against 26 strains of *Fusobacterium* spp., 0.03, > 128, 0.25, > 128, > 128 and 16 μ g/ml, respectively, against 25 strains of *Porphyromonas* spp., 0.06, 0.06, 4, > 128, > 128 and 32 μ g/ml, respectively, against 28 strains of *Prevotella* spp., 16, 4, 32, > 128, >

128 and 16 μ g/ml, respectively, against 31 strains of Clostridium spp., 0.03, 0.06, > 128, > 128, 8 and 0.5 μ g/ml, respectively, against 30 strains of *P. acnes*, and 1, > 128, 2, > 128, > 128 and 64 μ g/ml, respectively, against 28 strains of anaerobic Gram-positive cocci. Overall, the MIC₉₀ values against all 226 anaerobic isolates studied were 2 μ g/ml for retapamulin and clindamycin, > 128 μ g/ml for metronidazole, mupirocin and neomycin, and 32 μ g/ml for tetracycline (21).

The in vitro activities of retapamulin, mupirocin and moxifloxacin were tested against 974 mostly recent bacterial isolates. The MIC₉₀ values determined against all Staphylococcus strains combined (n=331) were 0.12, 1 and > 0.5 µg/ml for retapamulin, mupirocin and moxifloxacin, respectively. Among the 102 strains of coagulase-negative staphylococci tested, the MIC₉₀ values were 0.12, > 32 and > 0.5 μ g/ml for retapamulin, mupirocin and moxifloxacin, respectively, with similar levels of activity being seen against oxacillin-resistant and -susceptible strains. When tested against 229 strains of S. aureus, the agents yielded MIC₉₀ values of 0.12, 0.5 and > 0.5 μ g/ml, respectively. Among the subset of 109 oxacillin-resistant strains of S. aureus, the MIC_{q0} values were 0.12, 2 and > 0.5 μg/ml, respectively, and among the 20 vancomycin-intermediate strains of S. aureus, the respective MIC_{90} values were 0.12, 1 and > 0.5 μ g/ml. Testing against a total of 390 strains of streptococci revealed MIC_{on} values of 0.06, 2 and 0.25 μg/ml for retapamulin, mupirocin and moxifloxacin, respectively. The MIC_{on} values against all strains of *Enterococcus* spp. combined (n=102) were 128, > 32 and > 0.5 μ g/ml for retapamulin, mupirocin and moxifloxacin, respectively. Retapamulin demonstrated little activity against Gramnegative bacilli (n=151), with an MIC_{90} of > 512 μ g/ml; the corresponding MIC₉₀ values for mupirocin and moxifloxacin were > 32 and $> 0.5 \mu g/ml$ (22).

Retapamulin has a low potential to select for resistant mutants in S. pyogenes, according to a study that assessed the resistance generated against a number of antibacterial agents in multiple- and single-passage testing. In multiple-passage studies, retapamulin exhibited lower MIC values against S. pyogenes than the other agents tested (i.e., mupirocin, cefalexin, erythromycin, linezolid, vancomycin and quinupristin/dalfopristin). In 3 of the 10 isolates tested, retapamulin selected clones after 26-48 days. The MIC values were 0.125-0.25 μg/ml for these selected clones, as compared to MIC values of 0.016-0.03 µg/ml for the parents (representing an 8-fold increase). Even prolonged 50-day incubation with retapamulin did not raise the MIC values of these clones. In the single-passage studies, the resistance frequency against S. pyogenes was found to be lower for retapamulin that for the other agents tested. One multiple-passage clone had an L3 mutation (N149K) and was associated with an 8-fold increase in retapamulin MIC after 48 days; however, none of the 7 randomly selected single-passage clones exhibited altered L3 protein (23).

Another study determined the potential of retapamulin to select for resistant mutants in 12 strains of S. aureus,

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including 1 mupirocin-resistant and 1 fusidic acid-resistant strain, as well as 2 vancomycin-intermediate and 3 vancomycin-resistant strains. Mupirocin, fusidic acid, cefalexin, erythromycin, linezolid, vancomycin and quin-upristin/dalfopristin were used as comparators. After 14-20 days, retapamulin yielded resistant clones in all 12 strains, although retapamulin had the lowest MIC values of all the agents tested in the multiple-step resistance study. The MIC values for retapamulin-selected resistant clones were 8-32-fold higher that those for parents, but none of the MICs reached values above 2 μ g/ml. In single-passage resistance studies, the frequency of spontaneous resistance to retapamulin was lower than for all other agents tested except linezolid (24).

In a study to determine the postantibiotic effects (PAEs) of retapamulin and mupirocin, clinical isolates of S. aureus, S. pyogenes, S. pneumoniae and H. influenzae were incubated in the presence of 4 x MIC concentrations of each of the drugs for 2 h. The PAEs against S. aureus (3 strains) were found to be 3.1-3.4 h for retapamulin (MIC = $0.06-0.125 \mu g/ml$) and 2.2-2.9 h for mupirocin (MIC = $0.25-0.50 \mu g/ml$). When tested against S. pyogenes (4 strains), retapamulin showed a PAE of 3.5-4.2 h (MIC = $0.008-0.016 \,\mu\text{g/ml}$) and mupirocin had a PAE of 2.2-3.2 h (MIC = $0.06-0.125 \mu g/ml$). The PAEs against S. pneumoniae (2 strains) were 3.7-4.7 h for retapamulin (MIC = $0.03-0.06 \mu g/ml$) and 1.3-2.2 h for mupirocin (MIC = $0.125-2.00 \mu g/ml$). Finally, the results of testing against H. influenzae (2 strains) yielded PAEs of 3.3-5.2 h for retapamulin (MIC = 0.25-1.00 μ g/ml) and 2.1-3.5 h for mupirocin (MIC = $0.06 \mu g/ml$) (25).

Retapamulin showed excellent *in vitro* activity against 109 methicillin-resistant *S. aureus* isolates obtained from skin specimens collected from patients enrolled in phase III clinical trials. Results from this study demonstrated that retapamulin was active against all methicillin-resistant *S. aureus* isolates tested, independent of SCC*mec* (staphylococcal casette chromosome *mec*) type or the presence of PVL (Panton-Valentine leukocidin) genes. For 67 PVL-positive, SCC*mec* type IV isolates, the retapamulin MIC $_{50}$ and MIC $_{90}$ values were both 0.12 μ g/ml. When tested against 14 PVL-negative, SCC*mec* type IV isolates, retapamulin yielded MIC $_{50}$ and MIC $_{90}$ values of 0.06 and 0.12 μ g/ml, respectively. Against 28 PVL-negative, SCC*mec* type II, III or IIIA isolates, the retapamulin MIC $_{50}$ and MIC $_{90}$ values were 0.06 and 0.12 μ g/ml, respectively (26).

An *in vivo* study tested the efficacy of topical retapamulin against resistant strains of *S. aureus* in a murine wound infection model. The antibacterial efficacy of retapamulin (1% w/w b.i.d.) was compared to that of mupirocin (2% w/w t.i.d.) and fusidic acid (2% w/w t.i.d.). The retapamulin MIC values for the *S. aureus* strains tested were 0.12 µg/ml for F306 (methicillin- and azithromycin-resistant) and X32717 (methicillin-, mupirocin-, azithromycin- and levofloxacin-resistant), 0.06 µg/ml for T63256 (methicillin-, mupirocin-, azithromycin- and levofloxacin-resistant) and 1080 (mupirocin- and azithromycin- resistant), and 0.03 µg/ml for S5112 (mupirocin-, azithromycin- and levofloxacin- and levofloxacin-

resistant). Retapamulin also displayed equivalent or superior efficacy to the other agents in reducing bacterial counts in this infection model (27).

Drug Interactions

A randomized, open-label, crossover phase I study assessed the effects of ketoconazole on the pharmacokinetics of retapamulin in 28 healthy adult males. Volunteers were exposed for 24 h to 1% retapamulin ointment applied postabrasion to 50 cm² of skin either alone or on day 4 of oral ketoconazole treatment (200 mg b.i.d.). The mean retapamulin $\mathrm{AUC}_{\text{0-24h}}$ was 98.73 ng.h/ml in subjects receiving ketoconazole and 54.96 ng.h/ml in volunteers not receiving ketoconazole. The mean retapamulin C_{max} was 7.63 ng/ml in subjects receiving ketoconazole and 4.27 ng/ml in those not receiving ketoconazole. Oral ketoconazole thus induced an 81% increase in both the ${\rm AUC_{0-24h}}$ and ${\rm C_{max}}$ of retapamulin, although it did not affect the ${\rm t_{max}}$. Only infrequent and mild adverse events were observed. One subject withdrew early due to elevated serum creatinine levels during ketoconazole treatment and another subject withdrew due to a positive urine drug screen. No relationship was noted between cardiac Q-T interval and retapamulin plasma concentrations. The increased systemic retapamulin exposure observed with the co-administration of oral ketoconazole was not considered to be clinically relevant. Based on these results, the authors concluded that oral cytochrome P-450 CYP3A4 and P-glycoprotein inhibitors such as ketoconazole can be safely co-administered with topical retapamulin without the need for dose adjustment (28).

Clinical Studies

Retapamulin ointment is being tested in two phase III clinical studies for the topical treatment of impetigo (4-6).

Source

GlaxoSmithKline plc. (GB).

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